

Neurofilament Light Chain Levels Are Predictive of Clinical Conversion in Radiologically Isolated Syndrome

Manon Rival, MD,* Eric Thouvenot, MD, PhD,* Lucile Du Trieu de Terdonck, PhD, Sabine Laurent-Chabalier, PhD, Christophe Demattei, PhD, Ugur Uygunoglu, MD, Giovanni Castelnovo, MD, Mikael Cohen, MD, PhD, Darin T. Okuda, MD, PhD, Orhun H. Kantarci, MD, PhD, Daniel Pelletier, MD, Christina Azevedo, MD, PhD, Philippe Marin, PhD, Sylvain Lehmann, MD, PhD, Aksel Siva, MD, PhD, Thibault Mura, MD, PhD, and Christine Lebrun-Frenay, MD, PhD, on behalf of SFSEP and RISC

Correspondence
Prof. Thouvenot
eric.thouvenot@chu-nimes.fr

Neurol Neuroimmunol Neuroinflamm 2023;10:e200044. doi:10.1212/NXI.000000000200044

Abstract

Background and Objectives

To evaluate the predictive value of serum neurofilament light chain (sNfL) and CSF NfL (cNfL) in patients with radiologically isolated syndrome (RIS) for evidence of disease activity (EDA) and clinical conversion (CC).

Methods

sNfL and cNfL were measured at RIS diagnosis by single-molecule array (Simoa). The risk of EDA and CC according to sNfL and cNfL was evaluated using the Kaplan-Meier analysis and multivariate Cox regression models including age, spinal cord (SC) or infratentorial lesions, oligoclonal bands, CSF chitinase 3–like protein 1, and CSF white blood cells.

Results

Sixty-one patients with RIS were included. At diagnosis, sNfL and cNfL were correlated (Spearman $r = 0.78$, $p < 0.001$). During follow-up, 47 patients with RIS showed EDA and 36 patients showed CC (median time 12.6 months, 1–86). When compared with low levels, medium and high cNfL (>260 pg/mL) and sNfL (>5.0 pg/mL) levels were predictive of EDA (log rank, $p < 0.01$ and $p = 0.02$, respectively). Medium-high cNfL levels were predictive of CC (log rank, $p < 0.01$). In Cox regression models, cNfL and sNfL were independent factors of EDA, while SC lesions, cNfL, and sNfL were independent factors of CC.


Discussion

cNfL >260 pg/mL and sNfL >5.0 pg/mL at diagnosis are independent predictive factors of EDA and CC in RIS. Although cNfL predicts disease activity better, sNfL is more accessible than cNfL and can be considered when a lumbar puncture is not performed.

Classification of Evidence

This study provides Class II evidence that in people with radiologic isolated syndrome (RIS), initial serum and CSF NfL levels are associated with subsequent evidence of disease activity or clinical conversion.

MORE ONLINE

 **Class of Evidence**
Criteria for rating therapeutic and diagnostic studies
[NPub.org/coe](https://www.npub.org/coe)

*These authors contributed equally to the work as first authors.

From the Department of Neurology (M.R., E.T., G.C.), Nîmes University Hospital Center, Univ. Montpellier; Functional Genomics Institute (M.R., E.T., L.D.T.T., P.M.), Univ. Montpellier, CNRS, INSERM; Department of Biostatistics (S.L.-C., C.D., T.M.), Clinical Epidemiology, Public Health and Innovation in Methodology (BESPIM), Nîmes University Hospital Center, Univ. Montpellier, France; Department of Neurology (U.U., A.S.), Cerrahpasa School of Medicine, University of Istanbul, Turkey; Centre de Ressources et Compétences Sclérose En Plaques (CRCSEP) (M.C., C.L.-F.), CHU de Nice, Hôpital Pasteur 2, Université Côte d'Azur, UR2CA-URRIS, France; UT Southwestern Medical Center (D.T.O.), Dallas, TX; Mayo Clinic (O.H.K.), Rochester, MN; University of South California (D.P., C.A.), Los Angeles; and LBPC-PPC (S.L.), Univ. Montpellier, CHU Montpellier, INM, INSERM, France.

Go to [Neurology.org/NN](https://www.neurology.org/NN) for full disclosures. Funding information is provided at the end of the article.

The Article Processing Charge was funded by the CHU Nîmes.

SFSEP and RISC coinvestigators are listed in the appendix at the end of the article.

This is an open-access article distributed under the terms of the Creative Commons Attribution-Non Commercial-No Derivatives License 4.0 (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

Glossary

AUC = area under the curve; **CC** = clinical conversion; **CHI3L1** = CSF chitinase 3–like protein 1; **DIS** = dissemination in space; **EDA** = evidence of disease activity; **MS** = multiple sclerosis; **NEDA** = nonevidence of disease activity; **OCBs** = oligoclonal bands; **RIS** = radiologically isolated syndrome; **RRMS** = relapsing-remitting MS; **sNfL** = serum neurofilament light chain; **SC** = spinal cord.

Radiologically isolated syndrome (RIS) is a preclinical stage of multiple sclerosis (MS) characterized by MRI brain lesions typical of MS detected in patients with clinical conditions not suggestive of MS.¹ Given the absence of clinical features suggestive of MS, the gold standard for defining RIS is the specific dissemination in space (DIS) criteria (3 or 4 2005 DIS criteria for diagnosis of MS), which have not been updated since 2009 despite changes in MS criteria.^{1–3} Natural history studies suggest that the clinical course for patients with RIS is heterogeneous.^{4,5} Not all individuals will experience definable clinical symptoms attributable to inflammatory CNS demyelination and axonal damage. Identifying patients at high risk of clinical conversion and disability over time is mandatory for optimal clinical care. The risk for a future event may be stratified by key risk factors, but no biological biomarkers are used in clinical practice.^{4–6} Retrospective studies performed by an international working group, the Radiologically Isolated Syndrome Consortium (RISC), have demonstrated that one-third of patients with RIS converted to MS after 5 years and that another one-third experienced new brain lesions on follow-up scans.⁴ After 10 years, most patients with RIS convert to MS.⁵ Several factors have been associated with the risk of clinical conversion (CC) to MS in RIS: male sex, younger age, positive CSF for oligoclonal bands (OCBs), infratentorial (IT) lesions, spinal cord (SC) lesions on the index scan, and evidence of disease activity (EDA) defined by gadolinium-enhancing (Gd+) lesions on MRI follow-up.^{1,4,5,7,8} These prognostic factors have been confirmed on a large prospective cohort who had Gd+ lesions on the index scan.⁶

Several additional biomarkers have been explored for their specificity for MS. These include MRI markers such as the central vein sign that reflects perivenous inflammatory demyelination and can help differentiate MS from other white matter disorders.⁹ Another is the paramagnetic rim sign, which can be detected in most patients with RIS, suggesting the presence of subclinical chronically active demyelination at an early stage of the disease.¹⁰ Visual evoked potential anomalies and peripapillary retinal nerve fiber layer thickness measured by optical coherence tomography have also been associated with the risk of conversion to MS.^{7,11} Biological markers of MS in patients with RIS include CSF OCBs, high CSF neurofilament light chain (cNfL) levels, and IL-8 levels. By contrast, high CSF chitinase 3–like protein 1 (CHI3L1) and high CSF IL-17 levels are not associated with the risk of a clinical event.^{12–15} However, no peripheral biological marker providing additional prognostic value to epidemiologic, MRI, and CSF markers has so far been identified.¹⁶

CSF is a fluid of choice for characterizing biomarkers of brain disorders and has been widely investigated in MS at all stages of the disease.^{17,18} cNfL is a useful biomarker in several inflammatory and neurodegenerative CNS diseases.¹⁹ In MS, cNfL was mainly associated with EDA in relapsing-remitting MS (RRMS), clinically isolated syndromes (CIS), and RIS.^{12,20,21} Using an ultrasensitive single-molecule array (Simoa) technology, NfL levels can now be accurately determined in serum obtained from a blood sampling, avoiding invasive lumbar puncture. Serum NfL (sNfL) levels, investigated in MS at different stages of the disease, are associated with disease activity, treatment response, and long-term outcomes.^{22,23} They were also slightly increased in patients compared with those in controls many years before MS onset, suggesting that neuroaxonal damage already occurs during MS prodromal phase.²⁴

In this study, we explored the value of sNfL as a predictive biomarker of disease activity in patients with RIS. We evaluated the predictive value of sNfL and cNfL levels for EDA and CC in patients with RIS in the context of other prognostic factors.

Methods

This study followed the STARD 2015 (Standards for Reporting Diagnostic accuracy studies) reporting guideline.

Standard Protocol Approvals, Registrations, and Patient Consents

This study used biological samples previously withdrawn in a prospective study, which was approved by a local ethic committee with patient informed consent. Because this study retrospectively analyzed data from a cohort of patients, it was approved by the Institutional Review Board of Nîmes (IRB no. 22.01.09), and no participant consent was required, according to the French Law.²⁵

Patients

We selected a multicenter retrospective cohort of patients with untreated RIS from University Hospital Centers of Nice, Istanbul, and Nîmes on the basis of availability of CSF and/or serum samples. CSF samples from 50 patients and serum samples from 57 patients had been prospectively collected (biobank registered under no. 914066V2) between 2001 and 2018 after discovering unexpected demyelinating lesions fulfilling Okuda criteria,¹ processed locally and centralized for this study. Epidemiologic, MRI, and biological data had been collected from patients with RIS, and CC occurrence prospectively followed. Patients treated with disease-modifying

drugs during follow-up were not included in the cohort. Clinical conversion (CC) was defined as a first clinical event typical of MS (according to the 2017 revision of the McDonald criteria) in patients with RIS during follow-up.²⁶ Non-conversion (NC) was defined as the absence of a first clinical event specific to MS in patients with RIS during follow-up. Evidence of disease activity (EDA) was determined by CC and/or the presence of new T2 lesion(s) and/or new Gd+ lesion(s) on follow-up scans. Nonevidence of disease activity (NEDA) was defined as the absence of EDA during follow-up. In all cases, MRI data were retrieved from prospectively acquired information noted in the database of each center. There was no standardized MRI protocol, but all patients had 1.5T or 3.0T brain and spinal cord MRI studies. For all brain MRI studies, 3D FLAIR and 3D MPRAGE sequences were acquired. Standard CSF analyses from local laboratories were collected (red cell count, white blood cell [WBC] count, and CSF protein concentration). The presence of OCBs was also determined during biobanking as a routine test in each center by parallel isoelectric focusing of serum and CSF in combination with immunoblotting.

CSF and Serum Sample Collection

CSF samples were collected using polypropylene tubes at the end of lumbar punctures and centrifuged within 2 hours at 1,500 × g for 10 minutes according to the guidelines of the BioMS-*eu* consortium, except that sample transport and centrifugation were performed at 4°C instead of room temperature.²⁷ Aliquots (500 µL) were stored at -80°C in 1.5 mL tubes (Protein LoBind 0030108.116, Eppendorf) until use. Patients with traumatic lumbar punctures (>500 red cells/mm³) were excluded.

Serum samples were spun at the latest 2 hours after blood sampling at 1,500 × g for 10 minutes at room temperature and aliquots (500 µL) stored at -80°C in 1.5 mL tubes (Protein LoBind 0030108.116, Eppendorf) until use.

Measure of CSF and Serum Biomarkers

All serum and CSF samples were centralized at the Clinical Proteomics Platform of the Laboratory of Biochemistry—Clinical Proteomic in Montpellier. sNfL and cNfL concentrations were determined using commercial NF-Light assay (Quanterix, Billerica, MA) based on ultrasensitive Simoa technology, as previously described.²⁸ CSF CHI3L1 levels were measured by ELISA using the MicroVue YKL-40 EIA kit (Quidel Corporation, San Diego, CA), as already described.²⁹ All experiments were performed with a single batch of reagents by researchers blinded from any clinical information. There were no missing values for NfL or CHI3L1 measures.

Statistical Analysis

Statistical analyses were performed using the R software (v3.0.2) and SAS software, version 9.4 (SAS Institute, Cary, NC). The type I error rate was 0.05, and no correction for multiple testing was performed. Median NfL values were compared using the nonparametric Mann-Whitney test. Area under the curve (AUC) values were estimated from time-dependent ROC

curves for censored data using the inverse probability of censoring weighting technique of Uno method³⁰ for the occurrence of events at 2, 3, and 5 years. The time from RIS diagnosis to CC or EDA was analyzed using the Kaplan-Meier estimator and compared between groups using the log-rank test. Because log hazard of CC or EDA and the continuous value of cNfL and sNfL did not have a linear relationship, we chose to perform an analysis based on the tertiles of the distribution of these variables. These analyses did not show a risk gradient between the medium and high tertiles of cNfL and sNfL (see Results section Figure 2), indicating a possible threshold effect. We therefore continued our analyses by dichotomizing these variables on the value of the first tertile and thus grouping the medium and high tertiles. Age, sNfL (low vs medium-high), cNfL (low vs medium-high), CSF CHI3L1, CSF WBCs, OCBs, 3 or 4 2005 DIS criteria, and the presence Gd+ lesions, IT lesions, and SC lesions were tested in a univariate Cox model analysis, followed by a multivariate analysis when significant at a threshold of 0.2 in univariate analyses. Factors with more than 10% of missing data were not included in the multivariate models. Finally, we analyzed the additive value of sNfL and cNfL by comparing the predictive value of several predictive models based on the SC lesions and OCBs, with and without the addition of sNfL (low vs medium-high) or cNfL (low vs medium-high), analyzing the differences in Uno concordance statistic (for censored data) between the different models.³¹

Classification of Evidence

This study is based on an RIS cohort study with prospective data collection in local databases using EDMUS. RIS status was determined centrally according to the presence of 2009 RIS criteria without neurologic symptoms of MS, without knowledge of the diagnostic test results for OCBs, CSF CHI3L1 levels, CSF WBC count, sNfL levels, and cNfL levels. These markers were determined in more than 80% of the participants. Patients were prospectively followed up from RIS discovery with regular MRI and clinical evaluation.

Data Availability

Anonymized participant data that underlie the results reported in this article will be shared beginning 6 months and ending 12 months after article publication on reasonable request by qualified investigators.

Results

Patient Characteristics at Diagnosis

Sixty-one patients with RIS (75% females, mean age 37 years; SD 12) with a biocollection of serum (n = 57) and/or CSF (n = 50) samples were included in the study (Table 1). Of the processed samples, 47% had CSF OCBs, median WBC count was 2/mm³ (0–25), 23% had 4 2005 DIS criteria, 20% had at least 1 brain Gd+ lesion, and 29% had at least 1 infratentorial (IT) lesion. In a subgroup of 59 patients with spinal cord MRI, 47% had at least 1 SC lesion, none of which was enhanced after administration of gadolinium. Median (95% CI)

Table 1 Characteristics of the Total RIS Population and Subgroups According to Evidence of Disease Activity or Clinical Conversion During Follow-up

Patient characteristics	RIS n = 61	RIS-NC n = 25	RIS-CC n = 36	RIS-NEDA n = 14	RIS-EDA n = 47
Sex ratio (female %)	75	76	75	79	74
Mean age (y)	37	37	37	38	36
≥2 OCBs (%)	47	36	56	21	55
Median CSF WBC count (/mm ³)	2	1	2	1	2
Mean CSF WBC count (/mm ³)	3	2	4	2	4
Four 2005 DIS criteria, %	23	12	31	21	28
Presence of IT lesion(s), %	29	16	36	0	38
Presence of brain Gd+ lesion(s), %	20	28	14	21	19
Presence of SC lesion(s) (n = 59), %	47	17	67	21	53
Presence of SC Gd+ lesion(s) (n = 59), %	0	0	0	0	0
Median CSF CHI3L1 levels (ng/mL) (n = 50)	135	133	162	111	162
Median cNfL levels (pg/mL) (n = 50)	383	253	464	161	464
Median sNfL levels (pg/mL) (n = 57)	6.9	5.7	7.2	4.0	7.1

Abbreviations: CC = clinical conversion; CHI3L1 = chitinase 3-like protein 1; cNfL = CSF neurofilament light chain; DIS = dissemination in space; EDA = evidence of disease activity; Gd+ = gadolinium-enhancing; IT = infratentorial; NC = no clinical conversion; NEDA = nonevidence of disease activity; OCBs = oligoclonal bands; RIS = radiologically isolated syndrome; SC = spinal cord; sNfL = serum neurofilament light chain; WBCs = white blood cells

cNfL and sNfL levels were 383 (276; 569) pg/mL and 6.9 (5.3; 7.6) pg/mL, respectively, and median (95% CI) CSF CHI3L1 level was 135 (108; 183) ng/mL (Table 1).

In a subgroup of 46 patients (75%) with both sNfL and cNfL levels available, sNfL and cNfL levels were correlated (Spearman correlation coefficient $r = 0.780$, $p < 0.01$, Figure 1A). Neither sNfL ($r = 0.098$, $p = 0.47$) (Figure 1B) nor cNfL levels ($r = 0.020$, $p = 0.89$) were correlated with age (data not shown). sNfL levels were not significantly different between male and female patients, RIS patients with 3 or 4 2005 DIS criteria, and those with or without SC lesion (Figure 1, C and D). cNfL levels, but not sNfL levels, were significantly higher in RIS patients with positive OCBs than in those with negative OCBs ($p = 0.02$, Mann Whitney test, Figure 1, C and D).

Evolution of Patients With RIS During Follow-up

Patients were prospectively followed up from diagnosis of RIS for a median time of 23 months (2–228 months). During follow-up, 36 patients with RIS (59%) evolved to a CC with a median conversion time of 12.6 months (1–86 months), while 25 patients remained NC after a median follow-up of 78 months (7–228 months). Forty-seven patients with RIS (76%) showed EDA in a median delay of 11.1 months (1–86 months). The characteristics of the subgroups of patients regarding EDA vs NEDA and CC vs NC are summarized in Table 1. cNfL levels were higher in RIS patients with CC vs NC and EDA vs NEDA (464 pg/mL vs 253 pg/mL, $p = 0.5$, and 464 pg/mL vs 161 pg/mL, $p < 0.01$, respectively). sNfL levels were also higher

in RIS patients with EDA vs NEDA (7.1 pg/mL vs 4.0 pg/mL, $p < 0.01$), but not in RIS patients with CC compared with NC (7.2 vs 5.7 pg/mL, $p = 0.13$).

Association of cNfL and sNfL With Clinical Conversion and Evidence of Disease Activity

Using ROC curves, we determined time-dependent AUC [95% CI] values of sNfL and cNfL that best discriminate EDA vs NEDA and CC vs NC at 2, 3, and 5 years of follow-up (Table 2).

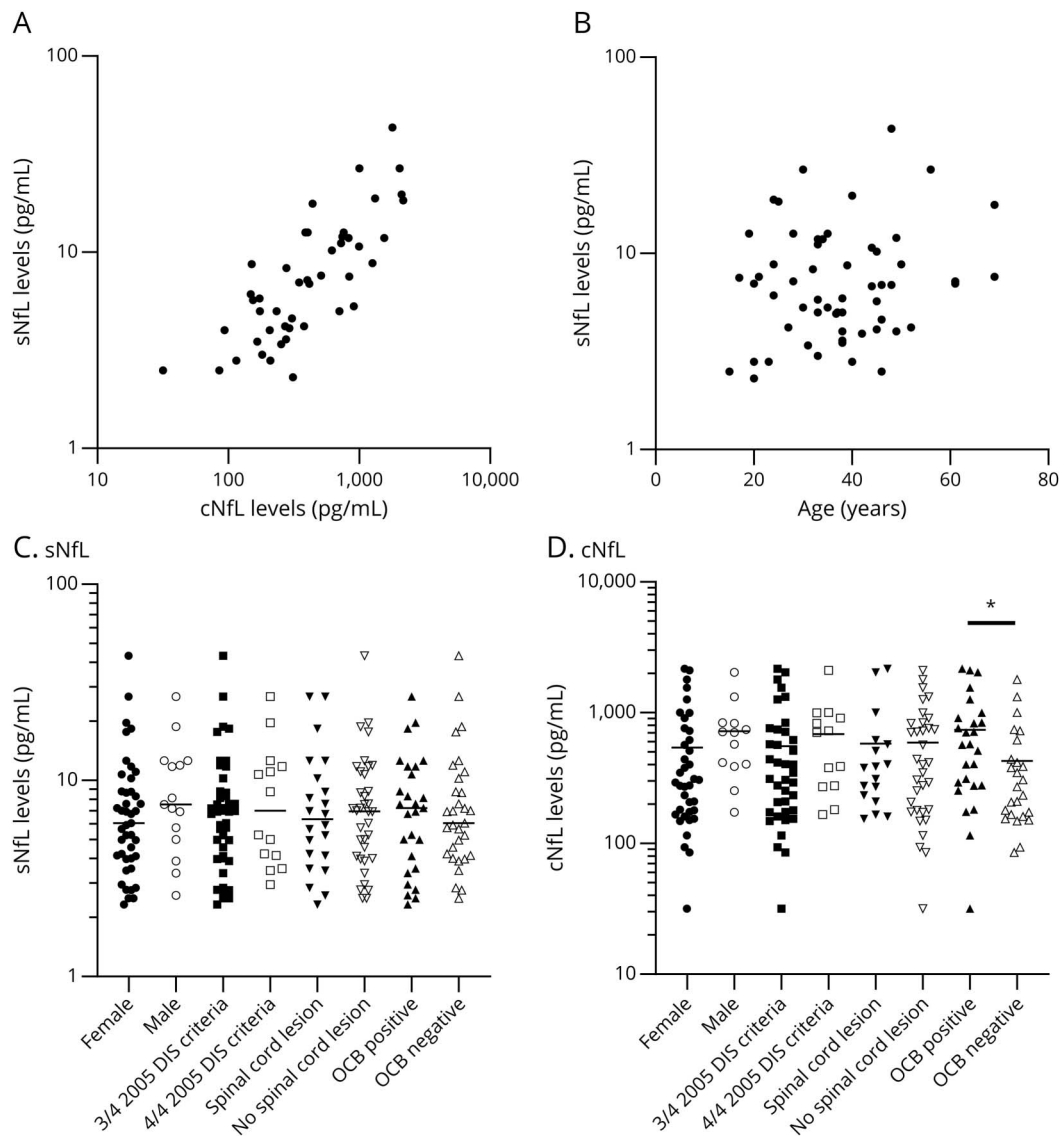
Best AUC values for CC vs NC were provided at 3 years (0.76 [0.59–0.92] for cNfL and 0.66 [0.47–0.86] for sNfL). Best AUC values for EDA vs NEDA were provided at 5 years (0.89 [0.76–1.00] for cNfL and 0.78 [0.54–1.00] for sNfL).

Comparison of cNfL and sNfL AUC [95% CI] values for CC and EDA at 3 years in a subgroup of 46 patients with both cNfL and sNfL levels available showed that cNfL has a more robust association profile with CC (0.77 [0.61; 0.92] and 0.62 [0.41; 0.84], respectively) and EDA (0.82 [0.65; 0.98] and 0.69 [0.49; 0.88], respectively) than sNfL, although not significantly.

Individual Factors Predicting Clinical Conversion and Evidence of Disease Activity

Using cutoff values defined as the first tertile of distribution as cutoff values (cNfL >260 and sNfL >5.0 pg/mL) and taking into account the time of follow-up, the Kaplan-Meier analysis revealed that the presence of IT lesion(s) and OCBs, medium-high cNfL, and medium-high sNfL are predictive of EDA (log rank, $p = 0.03$, $p = 0.03$, $p < 0.01$, and $p = 0.02$,

Figure 1 Association of sNfL and cNfL Levels With Different Clinical, MRI, and CSF Parameters



(A) Correlation between sNfL and cNfL levels. (B) sNfL and the age of patients with RIS are not correlated. (C) sNfL levels are not significantly different in subgroups determined by sex, number of 2005 DIS criteria, spinal cord lesion, or OCB positive. (D) cNfL levels are not significantly different in subgroups determined by sex, 2005 DIS criteria, or spinal cord lesion. They are significantly elevated in RIS patients with OCB compared with those in others ($*p = 0.02$, Mann-Whitney test). cNfL = CSF neurofilament light chain; DIS = dissemination in space; NfL = neurofilament light chain; OCBs = oligoclonal bands; RIS = radiologically isolated syndrome; sNfL = serum neurofilament light chain.

respectively, Figure 2, A and C, Figure 3, A and C). It also indicated that the presence of SC lesion(s), medium-high cNfL, and elevated WBCs ($\geq 5/\text{mm}^3$) are predictive of CC (log rank, $p < 0.01$, $p < 0.01$, and $p = 0.04$, respectively, Figure 2, B and D, Figure 3B), while medium-high sNfL was not (log rank, $p = 0.08$, Figure 3D). Age, CSF CHI3L1, number of 2005 DIS criteria, and the presence of Gd+ lesions on the index MRI were not significantly associated with EDA nor CC (data not shown).

Cox univariate analysis of age, sNfL, cNfL, CSF CHI3L1, CSF WBCs, OCBs, number of 2005 DIS criteria, Gd+ lesions, IT lesions, and SC lesions on the index MRI revealed that

medium-high cNfL and the presence of SC lesions were predictive of CC, while medium-high cNfL, medium-high sNfL, the presence of OCBs, and IT lesions were predictive of EDA (Table 3).

Multivariate Analysis of Prognostic Factors in RIS

cNfL and sNfL being highly correlated, we built 2 multivariate Cox regression models to identify EDA and CC prognostic factors in patients with RIS (Table 3).

In the multivariate model analyzing cNfL, medium-high cNfL was an independent predictive factor of EDA (HR = 8.0, $p <$

Table 2 Sensitivity, Specificity, and Area Under the Curve of sNfL and cNfL (Using First Tertile as Threshold for Positivity) for Evidence of Clinical Activity and Clinical Conversion at 2, 3, and 5 Years, Determined by Time-Dependent ROC Curves

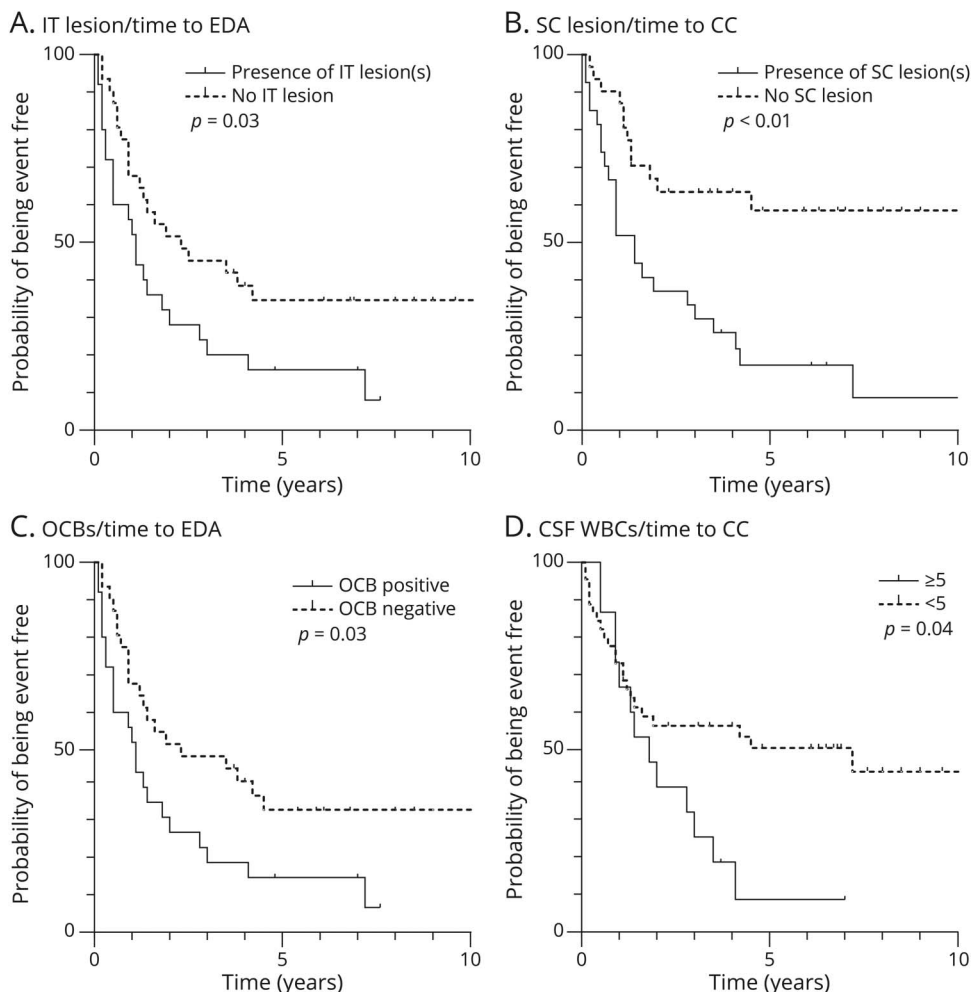
	2 y			3 y			5 y		
	Se (%)	Sp (%)	AUC (95% CI)	Se (%)	Sp (%)	AUC (95% CI)	Se (%)	Sp (%)	AUC (95% CI)
cNfL									
CC	83.3	62.5	0.71 (0.53; 0.88)	88.4	66.7	0.76 (0.59; 0.92)	85.5	58.3	0.68 (0.45; 0.92)
EDA	82.1	78.9	0.79 (0.62; 0.97)	87.1	75.0	0.81 (0.66; 0.96)	85.9	87.5	0.89 (0.76; 1.00)
sNfL									
CC	73.1	60.7	0.64 (0.47; 0.82)	67.7	68.0	0.66 (0.47; 0.86)	87.2	50.0	0.62 (0.42; 0.82)
EDA	67.7	69.6	0.67 (0.49; 0.85)	73.5	75.0	0.72 (0.55; 0.90)	87.3	66.7	0.78 (0.54; 1.00)

Abbreviations: AUC = area under the curve; CC = clinical conversion; cNfL = CSF neurofilament light chain; EDA = evidence of disease activity; Se = sensitivity; sNfL = serum neurofilament light chain; Sp = specificity.

0.01). By contrast, medium-high cNfL and the presence of SC lesion(s) were independent factors of CC (HR = 4.8, $p < 0.01$ and HR = 2.7, $p = 0.03$, respectively, Table 3).

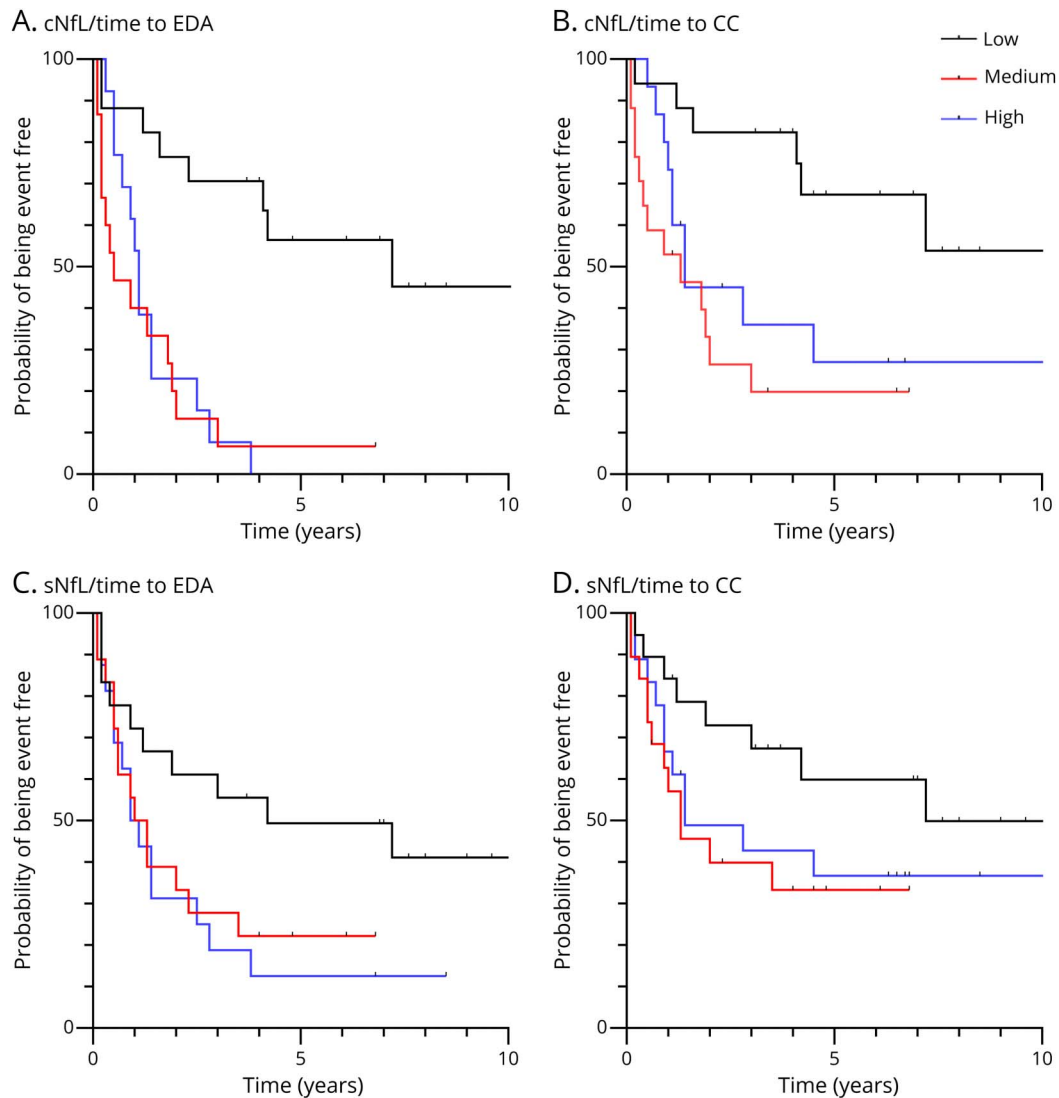
In the multivariate model analyzing sNfL, medium-high sNfL and SC lesions were independent predictive factors of EDA (HR = 3.0, $p = 0.01$ and HR = 2.2, $p = 0.03$, respectively),

Figure 2 Survival Curves for Evidence of Clinical Activity and Clinical Conversion According to Brain MRI and OCBs



Kaplan-Meier estimates of the probability of EDA and CC during follow-up are depicted as a function of the presence of IT lesions (A), SC lesions (B), the presence of OCBs (C), high levels of WBCs (D). CC = clinical conversion; EDA = evidence of disease activity; IT = infratentorial; OCBs = oligoclonal bands; SC = spinal cord; WBCs = white blood cells.

Figure 3 Survival Curves for Evidence of Clinical Activity and Clinical Conversion According to cNfL and sNfL



Kaplan-Meier estimates of the probability of EDA and CC during follow-up are depicted as a function of the presence of low (≤ 260 pg/mL), medium (260–710 pg/mL), or high (> 710 pg/mL) cNfL levels (A, B) and low (≤ 5.0 pg/mL), medium (5.0–8.5 pg/mL), or high (> 8.5 pg/mL) sNfL levels (C, D). CC = clinical conversion; cNfL = CSF neurofilament light chain; EDA = evidence of disease activity; sNfL = serum neurofilament light chain.

while medium-high sNfL, the presence of SC lesion(s), and OCBs were independent factors of CC (HR = 2.8, $p = 0.02$, HR = 3.3, $p < 0.01$, and HR = 2.36, $p = 0.02$, respectively) (Table 3). Especially, 84% of patients with RIS with medium-high sNfL levels, SC lesions, and OCBs had a CC at 3 years, compared with 34% in other patients with RIS.

To analyze the additive value of NfL to SC lesions and OCBs, we further compared the AUC values of SC lesions + OCBs and SC lesions + OCBs + NfL for EDA and CC at 3 years (Table 4). All AUCs values were improved when adding sNfL or cNfL to SC lesions and OCBs for the prediction of EDA and CC, although it was statistically significant only for cNfL and EDA at 3 years (AUC = 0.65 [0.43; 0.86] for SC lesions + OCBs, vs AUC = 0.85 [0.67; 1.00] for SC lesions + OCBs + cNfL, $p = 0.01$ for AUC comparison).

Classification of Evidence

This study provides Class II evidence that in people with RIS, initial serum and CSF NfL levels are associated with subsequent evidence of disease activity or clinical conversion.

Discussion

We compared the value of NfL levels in serum and CSF samples from patients with RIS in the context of clinical risk. We demonstrated that sNfL and cNfL levels were highly correlated and constituted independent predictors of CC. Our results align with the most extensive RIS studies that described the importance of SC lesions for RIS prognosis, also identified here as an independent predictor of CC in the multivariate Cox model.⁴⁻⁶ These factors reflect the risk of CC during follow-up. They could be considered to counsel patients with RIS during

Table 3 Analysis of Baseline Characteristics Predictive of Evidence of Disease Activity or Clinical Conversion During Follow-up Using Multivariate Cox Models

Variable	Univariate analysis			Variable	Multivariate analysis			
	HR	95% CI	p Value		Complete model	HR	95% CI	p Value
Time to CC								
cNFL ^a	3.9	1.6; 9.8	<0.01	cNFL (n = 50)	cNFL ^a	4.8	1.8; 12.8	<0.01
sNFL ^b	2.1	0.4; 4.5	0.07		SC	2.7	1.1; 6.7	0.03
SC	3.3	1.6; 6.6	<0.01		OCBs	1.8	0.8; 3.8	0.13
OCBs	1.8	0.9; 3.5	0.08		Gd+	0.6	0.2; 1.8	0.33
Gd+	0.4	0.2; 1.2	0.10		2005 DIS	0.7	0.3; 1.7	0.47
2005 DIS	1.6	0.8; 3.3	0.18	sNFL (n = 57)	sNFL ^b	2.8	1.2; 6.6	0.02
WBCs ^c	1.9	1.0; 3.9	0.07		SC	3.3	1.4; 7.7	<0.01
IT	1.5	0.7; 2.9	0.28		OCBs	2.4	1.1; 4.9	0.02
Age ^d	1.0	0.5; 1.9	0.89		Gd+	0.4	0.1; 1.7	0.22
CHI3L1 ^e	1.5	0.8; 3.0	0.213		2005 DIS	1.3	0.6; 2.7	0.55
Time to EDA								
cNFL ^a	5.9	2.5; 13.8	<0.01	cNFL (n = 50)	cNFL ^a	8.0	2.8; 22.7	<0.01
sNFL ^b	2.2	1.1; 4.5	0.02		OCBs	1.3	0.6; 2.7	0.43
OCBs	2.1	1.2; 3.7	0.01		SC	1.8	0.9; 3.9	0.12
SC	1.6	0.9; 2.9	0.12		CHI3L1 ^e	1.0	0.5; 2.1	0.99
CHI3L1 ^e	1.6	0.9; 3.0	0.12		IT	1.0	0.5; 2.1	0.96
IT	2.1	1.2; 3.9	0.01	sNFL (n = 57)	sNFL ^b	3.0	1.3; 6.8	0.01
2005 DIS	1.5	0.8; 2.9	0.19		OCBs	1.7	0.8; 3.8	0.17
Gd+	0.8	0.4; 1.7	0.55		SC	2.2	1.1; 3.8	0.03
Age ^d	0.9	0.5; 1.7	0.78		CHI3L1 ^e	1.1	0.5; 2.4	0.75
WBCs ^c	1.0	0.5; 2.0	0.94		IT	1.3	0.6; 3.0	0.45

Abbreviations: 2005 DIS = presence of four 2005 DIS criteria; CHI3L1 = chitinase 3-like protein 1; cNFL = CSF neurofilament light chain; EDA = evidence of disease activity; Gd+ = presence of brain gadolinium-enhanced lesion(s); HR = hazard ratio; IT = infratentorial; LCC = clinical conversion; NfL = neurofilament light chain; OCBs = oligoclonal bands; SC = presence of spinal cord lesion(s); sNFL = serum neurofilament light chain; sNFL = serum NfL; WBCs = white blood cells.

Items with a p value <0.2 were included in the multivariate Cox regression models and those with a p value <0.05 are in bold.

^a cNFL >260 pg/mL.

^b sNFL >5.0 pg/mL.

^c WBCs ≥5/mm³.

^d Age older than 37 years.

^e CSF CHI3L1 > 135 ng/mL.

diagnosis and to organize the clinical and MRI surveillance to detect disease activity. Indeed, EDA is a classical endpoint in the follow-up of patients with MS, but predictive factors of EDA have never been investigated in patients with RIS. Identifying OCBs and SC lesions as predictors of EDA also suggests that these factors provide complementary information to sNFL and cNFL that remain independent predictive factors in both cases (EDA and CC). Moreover, sNFL or cNFL provide additive value to SC lesions and OCBs to discriminate RIS patients with CC or EDA from clinically and/or radiologically stable RIS patients,

especially for cNFL and EDA (Table 4). As a matter of fact, NfL are released from axons and reflect neuroaxonal injury in MS. NfL seems to be more specifically associated with focal inflammation (active plaques and relapses) than to the neurodegenerative process associated with disease progression.³²

Recently, improved detection of sNFL based on ultrasensitive ELISA kits or Simoa allowed to reliably measure sNFL concentrations that are well correlated to that of CSF in patients with MS.³³ The high sensitivity of this test, illustrated by the

Table 4 Additive Value of sNfL and cNfL to SC Lesions and OCBs to Discriminate CC and EDA From Clinically and/or Radiologically Stable RIS Patients at 3 Years

Endpoint	Combinations of variables	Se (%)	Sp (%)	AUC (95% CI)	AUC comparison p Value
cNfL (n = 50)					
CC	SC lesion + OCBs	40.0	89.0	0.70 (0.55; 0.86)	0.13
	SC lesion + OCBs + cNfL	80.0	81.0	0.85 (0.71; 0.99)	
EDA	SC lesion + OCBs	66.7	61.0	0.65 (0.43; 0.86)	0.01
	SC lesion + OCBs + cNfL	93.3	77.4	0.85 (0.67; 1.00)	
sNfL (n = 57)					
CC	SC lesion + OCBs	70.8	68.0	0.77 (0.66; 0.88)	0.46
	SC lesion + OCBs + sNfL	66.7	78.5	0.79 (0.64; 0.94)	
EDA	SC lesion + OCBs	47.3	84.8	0.71 (0.56; 0.86)	0.65
	SC lesion + OCBs + sNfL	78.9	69.7	0.77 (0.61; 0.93)	

Items with a *p* value <0.05 are in bold.

AUC = area under the curve (Uno Concordance Statistic); CC = clinical conversion; cNfL = CSF neurofilament light chain >260 pg/mL; EDA = evidence of disease activity; OCBs = positive oligoclonal bands; RIS = radiologically isolated syndrome; SC lesion = presence of spinal cord lesion(s); Se = sensitivity; sNfL = serum neurofilament light chain >5.0 pg/mL; Sp = specificity.

absence of missing values, allows accurate measures of sNfL in serum. This study provides the first measurement of sNfL in patients with RIS using the NF-light assay. Using this assay, the median sNfL (6.9 pg/mL) in patients with RIS was lower than that observed by others in patients with presymptomatic MS (16.7 pg/mL) and patients with CIS (17.0 pg/mL) using the same antibody adapted to Simoa homebrew kits.^{24,34} However, our results corroborate sNfL median levels measured in our previous study using the NF-light assay in patients with MS (9.4 pg/mL)³⁵ and those recently found in patients with early RRMS (10.1 pg/mL) using the same assay.³⁶

Compared with the electrochemiluminescence-based method or ELISA, the high accuracy of the Simoa assay, even at lower values, allows sNfL to strongly correlate with cNfL.³⁷ This probably contributed to the performance of sNfL as a prognostic biomarker of clinical evolution in patients with RIS in our study. However, cNfL showed an overall more robust association profile with CC and EDA, suggesting that cNfL better reflects disease activity and predicts the evolution of patients with RIS than sNfL.³⁴ This might be the consequence of differences among individuals in the kinetics of neurofilament protein release from neurons and trafficking between the brain and blood compartments.³² Nevertheless, blood sampling is less invasive, making sNfL an excellent alternative to cNfL for RIS prognosis when a lumbar puncture cannot be performed. More extensive validation studies are required before sNfL may replace cNfL.

The high accuracy of the Simoa assay probably enabled sNfL to overcome OCBs to predict CC. Even if this suggests that OCBs might no longer be required for the prognosis of CC in RIS patients with SC lesions, their detection provides a reasonable specificity for the differential diagnosis of RIS vs other conditions.

Contrasting with the observations of a previous study¹² showing that OCBs and cNfL were independent predictive factors of CC in patients with RIS, OCBs and sNfL were independent predictors of CC in our study. Including the presence of SC lesions, one of the most robust predictors of CC in our Cox regression analysis might explain why cNfL and SC lesions, but not OCBs, were independent predictors of CC in our study. This discrepancy might also reflect the different populations, time of follow-up, and assays used to measure NfL (ELISA vs Simoa) or the association between the presence of OCBs and cNfL levels that potentially minimize their prognostic value (Figure 1D). Nevertheless, we confirm that CSF CHI3L1 is not a predictor of EDA and CC in patients with RIS.^{12,14}

This study has some limitations given its retrospective nature. Incomplete availability of CSF and serum samples to determine cNfL, CSF CHI3L1, and sNfL led to missing data, lowering the power of the regression models, limited to 50 and 57 patients for the cNfL and the sNfL models, respectively (Table 3). Moreover, the small size and the small number of centers limit knowledge regarding how these data are translatable to other races and ethnicities with RIS. In addition, although our population was comparable with cohorts from previous published RIS studies (age, sex ratio, MRI, and CSF findings), the proportion of patients with RIS converting to MS was higher (50% converters at 36 months) in our cohort.^{4,5} This might be explained by the selection of patients with untreated RIS only. This also probably enhanced the statistical power of the study for identifying predictors of CC and constitutes a potential caveat for future confirmation studies with less active cohorts. As illustrated in the Kaplan-Meier analyses, cNfL and sNfL values did not

appear as proportionally correlated with CC and EDA risk in a linear relationship, with similar risks of conversion for medium values and high values of each marker, when compared with low values (Figure 2). This suggests that NfL increase above a threshold value of 260 pg/mL for cNfL and 5.0 pg/mL for sNfL (corresponding to the first tertile of cNfL and sNfL in our study) indicates active neuroaxonal injury predictive of EDA and CC, especially for cNfL (Table 3). Intriguingly, the cutoff value for cNfL to predict CC was lower using Simoa (260 pg/mL) than that previously determined by ELISA in patients with RIS (619 pg/mL).¹² It can be the consequence of different assays used to measure NfL (ELISA vs Simoa) and of diverse populations investigated, as already observed in previous studies dedicated to MS biomarkers, such as those examining CHI3L1 in patients with CIS.^{29,38} A threshold value determined using Simoa of 5.0 pg/mL could be chosen for prospective studies aiming at validating the performance of sNfL alone or in combination with SC lesions and OCBs to predict CC and EDA and at establishing an RIS conversion score combining these factors.

In conclusion, we showed that elevated cNfL and sNfL levels at diagnosis are both predictive factors of EDA and CC in RIS. Our study provides the first analysis of sNfL in an RIS cohort and offers Class I evidence that sNfL >5.0 pg/mL is an independent predictor of CC and EDA in patients with RIS. Although cNfL predicts disease activity better, sNfL is more accessible and can be considered to identify patients at higher risk of conversion to help clinicians for counseling and monitoring patients when a lumbar puncture is not performed.

Acknowledgment

The authors are most grateful to Sarah Kabani, Medical Writer at the B.E.S.P.I.M, Nîmes University Hospital, for revising and editing this article.

Study Funding

The study was funded by CHU de Nîmes and has also been supported by the University of Montpellier, the Centre National de la Recherche Scientifique (CNRS), the Institut National de la Santé et de la Recherche Médicale (INSERM), the Occitanie Region, the fondation pour l'aide à la recherche sur la Sclérose en Plaques (ARSEP Fondation), the Radiologically Isolated Syndrome Consortium (RISC), and the Société Francophone de la Sclérose en Plaques (SFSEP).

Disclosure

M. Rival has nothing to disclose. E. Thouvenot has received honoraria, travel grants, or research grants from the following pharmaceutical companies: Actelion, Biogen, Genzyme, Merck Serono, Novartis, Roche, and Teva pharma. L. Du Trieu de Terdonck has nothing to disclose. S. Laurent-Chabalier has nothing to disclose. C. Demattei has nothing to disclose. U. Uygunoglu has nothing to disclose. G. Castelnovo has nothing to disclose. M. Cohen received personal fees from Biogen, Merck, Novartis, Roche, Alexion, Ad

Scientiam, Teva, and Sanofi. D.T. Okuda received personal compensation for consulting and advisory services from Biogen, Celgene/Bristol Myers Squibb, EMD Serono, Genentech, Genzyme, Janssen Pharmaceuticals, Novartis, Osmotica Pharmaceuticals, RVL Pharmaceuticals, Inc., TG Therapeutics, and Viela Bio, Inc. and research support from Biogen and EMD Serono/Merck. Royalties received for intellectual property licensed by The Board of Regents of The University of Texas System. O.H. Kantarci has nothing to disclose. Daniel Pelletier has nothing to disclose. C. Azevedo has nothing to disclose. S. Lehmann has nothing to disclose. P. Marin has nothing to disclose. A. Siva has received honoraria for educational presentations internationally and at national meetings and symposia sponsored by Bayer-Schering AG; Merck-Serono; Teva-Turkey; Biogen Idec/Gen Pharma of Turkey and Genzyme and consultation fees or travel and registration coverage for attending several national or international congresses or symposia from Merck Serono, Novartis, Genzyme, and Roche. T. Mura has nothing to disclose. C. Lebrun-Frenay has participated in expert boards for Biogen, Novartis, Roche, and Genzyme in the last 5 years. Expert and Speaker honoraria are either declined or donated to the URRIS research unit, University Cote d'Azur, Nice, France. She did not receive any financial compensation for her participation in the scientific committee of the French MS Society, ARSEP, andECTRIMS apart from travel expenses. Go to Neurology.org/NN for full disclosures.

Publication History

Received by *Neurology: Neuroimmunology & Neuroinflammation* March 8, 2022. Accepted in final form August 29, 2022. Submitted and externally peer reviewed. The handling editor was Friedemann Paul, MD.

Appendix 1 Authors

Name	Location	Contribution
Manon Rival, MD	CHU Nîmes, France; IGF, Montpellier, France	Acquired and analyzed the data; revised the article for intellectual content
Eric Thouvenot, MD, PhD	CHU Nîmes, France; IGF, Montpellier, France	Design and conceptualized study; analyzed the data; and drafted the article for intellectual content
Lucile Du Trieu de Terdonck, PhD	IGF, Montpellier, France	Acquired and analyzed the data; revised the article for intellectual content
Sabine Laurent-Chabalier, PhD	CHU Nîmes, France	Interpreted the data; revised the article for intellectual content
Christophe Demattei, PhD	CHU Nîmes, France	Interpreted the data; revised the article for intellectual content
Ugur Uygunoglu, MD	University of Istanbul, Turkey	Significant role in the acquisition of data
Giovanni Castelnovo, MD	CHU Nîmes, France	Significant role in the acquisition of data; revised the article for intellectual content

Appendix 1 (continued)

Name	Location	Contribution
Mikael Cohen, MD, PhD	CHU Nice, France	Major role in the acquisition of data; revised the article for intellectual content
Darin T. Okuda, MD, PhD	UTSMC, Dallas, Texas	Interpreted the data; revised the article for intellectual content
Orhun H. Kantarci, MD, PhD	Mayo Clinic, Rochester, Minnesota	Interpreted the data; revised the article for intellectual content
Daniel Pelletier, MD	USC, Los Angeles, California	Interpreted the data; revised the article for intellectual content
Christina Azevedo, MD, PhD	USC, Los Angeles, California	Interpreted the data; revised the article for intellectual content
Philippe Marin, PhD	IGF, Montpellier, France	Interpreted the data; revised the article for intellectual content
Sylvain Lehmann, MD, PhD	LBPC-PPC, Montpellier, France	Major role in the acquisition of data; interpreted the data; and revised the article for intellectual content
Aksel Siva, MD, PhD	University of Istanbul, Turkey	Interpreted the data; revised the article for intellectual content
Thibault Mura, MD, PhD	CHU Nîmes, France	Analyzed the data; revised the article for intellectual content
Christine Lebrun-Frenay, MD, PhD	CHU Nice, France	Design and conceptualized study; analyzed the data; and revised the article for intellectual content

Appendix 2 Coinvestigators

Name	Location	Role	Contribution
Orhun Kantarci, MD	Mayo Clinic, Rochester, MN, USA	RISC Site Investigator	Acquisition of data
Aksel Siva, MD	Istanbul University Cerrahpasa School of Medicine, Istanbul, Turkey	RISC Site Investigator	Acquisition of data
Daniel Pelletier, MD	University of Southern California, Los Angeles, CA, USA	RISC Site Investigator	Acquisition of data
Darin Okuda, MD	University of Texas South Western Medical Center, Dallas, TX, USA	RISC Site Investigator	Acquisition of data
Christina Azevedo, MD	University of Southern California, Los Angeles, CA, USA	RISC Site Investigator	Acquisition of data
Naila Makhani, MD	Departments of Pediatrics and Neurology, Yale School of Medicine, New Haven, CT, USA	RISC Site Investigator	Acquisition of data
Christine Lebrun-Frenay, MD, PhD	Nice University Hospital, UR2CA, Nice, France	RISC Site Investigator	Acquisition of data
Mikael Cohen, MD, PhD	Nice University Hospital, UR2CA, Nice, France	RISC Site Investigator	Acquisition of data

Appendix 2 (continued)

Name	Location	Role	Contribution
Lydiane Mondot, MD	Nice University Hospital, UR2CA, Nice, France	RISC Site Investigator	Acquisition of data
Eric Thouvenot, MD, PhD	Nîmes University Hospital, Nîmes, France	RISC Site Investigator	Acquisition of data
Jonathan Ciron, MD	Toulouse University Hospital, Toulouse, France	RISC Site Investigator	Acquisition of data
Françoise Durand-Dubief, MD, PhD	Hospices Civils de Lyon, Lyon, France	RISC Site Investigator	Acquisition of data
Adullatif Alkhedr, MD	Amiens University Hospital, Amiens, France	SFSEP Site Investigator	Acquisition of data
Geraldine Androdiadis, MD	Hospices Civils de Lyon, Lyon, France	SFSEP Site Investigator	Acquisition of data
Bertrand Audoin, MD, PhD	Assistance Publique des Hôpitaux de Marseille, Marseille, France	SFSEP Site Investigator	Acquisition of data
Xavier Ayrignac, MD, PhD	Montpellier University Hospital, Montpellier, France	SFSEP Site Investigator	Acquisition of data
Caroline Bensa, MD	Fondation Rothschild, Paris, France	SFSEP Site Investigator	Acquisition of data
Eric Berger, MD	Besancon University Hospital, Besancon, France	SFSEP Site Investigator	Acquisition of data
Damien Biotti, MD	Toulouse University Hospital, Toulouse, France	SFSEP Site Investigator	Acquisition of data
Bertrand Bourre, MD, PhD	Rouen University Hospital, Rouen, France	SFSEP Site Investigator	Acquisition of data
Latine Boyer, MD	Neurologist, La Rochelle, France	SFSEP Site Investigator	Acquisition of data
Pierre Branger, MD	Caen University Hospital, Caen, France	SFSEP Site Investigator	Acquisition of data
Saskia Bresch, MD	Nice University Hospital, UR2CA, Nice, France	SFSEP Site Investigator	Acquisition of data
Bruno Brochet, MD, PhD	Bordeaux University Hospital, Bordeaux, France	SFSEP Site Investigator	Acquisition of data
Philippe Cabre, MD, PhD	Martinique University Hospital, Fort de France, France	SFSEP Site Investigator	Acquisition of data
Jean-Philippe Camdessanché, MD, PhD	Saint-Etienne University Hospital, Saint-Etienne, France	SFSEP Site Investigator	Acquisition of data
Clarisse Carra-Dalliere, MD	Montpellier University Hospital, Montpellier, France	SFSEP Site Investigator	Acquisition of data
Olivier Casez, MD	Grenoble-Alpes University Hospital, Grenoble, France	SFSEP Site Investigator	Acquisition of data
Giovanni Castelnovo, MD	Nîmes University Hospital, Nîmes, France	SFSEP Site Investigator	Acquisition of data
Jonathan Ciron, MD	Toulouse University Hospital, Toulouse, France	SFSEP Site Investigator	Acquisition of data

Continued

Appendix 2 (continued)

Name	Location	Role	Contribution
Pierre Clavelou, MD, PhD	Clermont-Ferrand University Hospital, Clermont-Ferrand, France	SFSEP Site Investigator	Acquisition of data
Mikael Cohen, MD, PhD	Nice University Hospital, UR2CA,, Nice, France	SFSEP Site Investigator	Acquisition of data
Nicolas Collongues, MD, PhD	Strasbourg University Hospital, Strasbourg, France	SFSEP Site Investigator	Acquisition of data
Alain Créange, MD, PhD	Assistance Publique des Hôpitaux de Paris, Henri Mondor Hospital, Paris, France	SFSEP Site Investigator	Acquisition of data
Jerome De Seze, MD, PhD	Strasbourg University Hospital, Strasbourg, France	SFSEP Site Investigator	Acquisition of data
Marc Debouverie, MD, PhD	Nancy University Hospital, Nancy, France	SFSEP Site Investigator	Acquisition of data
Gilles Defer, MD, PhD	Caen University Hospital, Caen, France	SFSEP Site Investigator	Acquisition of data
Nathalie Derache, MD	Caen University Hospital, Caen, France	SFSEP Site Investigator	Acquisition of data
Françoise Durand-Dubief, MD, PhD	Hospices Civils de Lyon, Lyon, France	SFSEP Site Investigator	Acquisition of data
Bertrand Fontaine, MD, PhD	Sorbonne University, Paris, France	SFSEP Site Investigator	Acquisition of data
Agnès Fromont, MD, PhD	Dijon University Hospital, Dijon, France	SFSEP Site Investigator	Acquisition of data
Claire Giannesini, MD	Assistance Publique des Hôpitaux de Paris, Saint-Antoine Hospital, Paris, France	SFSEP Site Investigator	Acquisition of data
Olivier Gout, MD, PhD	Fondation Rothschild, Paris, France	SFSEP Site Investigator	Acquisition of data
Deborah Grosset-Jeannin, MD	Neurologist, Annecy, France	SFSEP Site Investigator	Acquisition of data
Antoine Guegen, MD	Fondation Rothschild, Paris, France	SFSEP Site Investigator	Acquisition of data
Anne Marie Guennoc, CRA	Toulouse University Hospital, Tours, France	SFSEP Site Investigator	Acquisition of data
Patrick Hautecoeur, MD, PhD	Lille Catholic University, Lille, France	SFSEP Site Investigator	Acquisition of data
Olivier Heinzeff, MD	Poissy-Saint-Germain Hospital, Poissy, France	SFSEP Site Investigator	Acquisition of data
Iuliana Ionescu, MD	Hospices Civils de Lyon, Lyon, France	SFSEP Site Investigator	Acquisition of data
Pierre Labauge, MD, PhD	Montpellier University Hospital, Montpellier, France	SFSEP Site Investigator	Acquisition of data
David Laplaud, MD, PhD	Nantes University Hospital, Nantes, France	SFSEP Site Investigator	Acquisition of data

Appendix 2 (continued)

Name	Location	Role	Contribution
Emmanuelle Le Page, MD	Rennes University Hospital, Rennes, France	SFSEP Site Investigator	Acquisition of data
Christine Lebrun-Frénay, MD, PhD	Nice University Hospital, UR2CA,, Nice, France	SFSEP Site Investigator	Acquisition of data
Emmanuelle Leray, PhD	Ecole des Hautes Etudes en Santé Publique, Rennes, France	SFSEP Site Investigator	Acquisition of data
Celine Louapre, MD, PhD	Assistance Publique des Hôpitaux de Paris, Pitié-Salpêtrière Hospital, Paris, France	SFSEP Site Investigator	Acquisition of data
Catherine Lubetzki, MD, PhD	Assistance Publique des Hôpitaux de Paris, Pitié-Salpêtrière Hospital, Paris, France	SFSEP Site Investigator	Acquisition of data
Adil Maarouf, MD, PhD	Assistance Publique des Hôpitaux de Marseille, Marseille, France	SFSEP Site Investigator	Acquisition of data
Laurent Magy, MD, PhD	Limoges University Hospital, Limoges, France	SFSEP Site Investigator	Acquisition of data
Elisabeth Maillart, MD	Assistance Publique des Hôpitaux de Paris, Pitié-Salpêtrière Hospital, Paris, France	SFSEP Site Investigator	Acquisition of data
Guillaume Mathey, MD	Nancy University Hospital, Nancy , France	SFSEP Site Investigator	Acquisition of data
Laure Michel, MD, PhD	Rennes University Hospital, Rennes, France	SFSEP Site Investigator	Acquisition of data
Lydiane Mondot, MD	Nice University Hospital, UR2CA,, Nice, France	SFSEP Site Investigator	Acquisition of data
Thibault Moreau, MD, PhD	Dijon University Hospital, Dijon, France	SFSEP Site Investigator	Acquisition of data
Nathalie Morel, MD	Neurologist, Annecy, France	SFSEP Site Investigator	Acquisition of data
Jean-Christophe Ouallet, MD	Bordeaux University Hospital, Bordeaux, France	SFSEP Site Investigator	Acquisition of data
Olivier Outteryck, MD, PhD	Lille University Hospital, Lille, France	SFSEP Site Investigator	Acquisition of data
Caroline Papeix, MD	Assistance Publique des Hôpitaux de Paris, Pitié-Salpêtrière Hospital, Paris, France	SFSEP Site Investigator	Acquisition of data
Jean Pelletier, MD, PhD	Assistance Publique des Hôpitaux de Marseille, Marseille, France	SFSEP Site Investigator	Acquisition of data
Fatai Radji, MD	Neurologist, Agens, France	SFSEP Site Investigator	Acquisition of data
Aurélien Ruet, MD, PhD	Bordeaux University Hospital, Bordeaux, France	SFSEP Site Investigator	Acquisition of data
Bruno Stankoff, MD, PhD	Assistance Publique des Hôpitaux de Paris, Saint-Antoine Hospital, Paris, France	SFSEP Site Investigator	Acquisition of data

Appendix 2 (continued)

Name	Location	Role	Contribution
Laurent Suchet, MD	Neurologist, Marseille, France	SFSEP Site Investigator	Acquisition of data
Frédéric Taithe, MD	Clermont-Ferrand University Hospital, Clermont-Ferrand, France	SFSEP Site Investigator	Acquisition of data
Eric Thouvenot, MD, PhD	Nîmes University Hospital, Nîmes, France	SFSEP Site Investigator	Acquisition of data
Ayman Tourbah, MD, PhD	APHP IdF Ouest, Paris Saclay University, Paris, France	SFSEP Site Investigator	Acquisition of data
Aurelian Ungureanu, MD	Neurologist, Annecy, France	SFSEP Site Investigator	Acquisition of data
Patrick Vermersch, MD, PhD	Lille University Hospital, Lille, France	SFSEP Site Investigator	Acquisition of data
Sandrine Wiertlewski, MD	Nantes University Hospital, Nantes, France	SFSEP Site Investigator	Acquisition of data
Helene Zephir, MD, PhD	Lille University Hospital, Lille, France	SFSEP Site Investigator	Acquisition of data

References

- Okuda DT, Mowry EM, Beheshtian A, et al. Incidental MRI anomalies suggestive of multiple sclerosis: the radiologically isolated syndrome. *Neurology*. 2009;72(9):800-805.
- Polman CH, Reingold SC, Banwell B, et al. Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald criteria. *Ann Neurol*. 2011;69(2):292-302.
- Polman CH, Reingold SC, Edan G, et al. Diagnostic criteria for multiple sclerosis: 2005 revisions to the "McDonald criteria". *Ann Neurol*. 2005;58(6):840-846.
- Okuda DT, Siva A, Kantarci O, et al; Radiologically Isolated Syndrome Consortium RISC; Club Francophone de la Sclérose en Plaques CFSEP. Radiologically isolated syndrome: 5-year risk for an initial clinical event. *PLoS One*. 2014;9(3):e90509.
- Lebrun-Frenay C, Kantarci O, Siva A, Sormani MP, Pelletier D, Okuda DT; 10-Year RISC Study Group on behalf of SFSEP, OFSEP. Radiologically isolated syndrome: 10-year risk estimate of a clinical event. *Ann Neurol*. 2020;88(2):407-417.
- Lebrun-Frenay C, Rollot F, Mondot L, et al; RISC, SFSEP, and OFSEP Investigators. Risk factors and time to clinical symptoms of multiple sclerosis among patients with radiologically isolated syndrome. *JAMA Netw Open*. 2021;4(10):e2128271.
- Lebrun C, Bensa C, Debouverie M, et al; Club Francophone de la Sclérose en Plaques. Association between clinical conversion to multiple sclerosis in radiologically isolated syndrome and magnetic resonance imaging, cerebrospinal fluid, and visual evoked potential: follow-up of 70 patients. *Arch Neurol*. 2009;66(7):841-846.
- Lebrun C, Bensa C, Debouverie M, et al; CFSEP. Unexpected multiple sclerosis: follow-up of 30 patients with magnetic resonance imaging and clinical conversion profile. *J Neurol Neurosurg Psychiatry*. 2008;79(2):195-198.
- Suthiphosuwat S, Sati P, Guenette M, et al. The central vein sign in radiologically isolated syndrome. *AJNR Am J Neuroradiol*. 2019;40(5):776-783.
- Suthiphosuwat S, Sati P, Absinta M, et al. Paramagnetic rim sign in radiologically isolated syndrome. *JAMA Neurol*. 2020;77(5):653-655.
- Aly L, Havla J, Lepenietter G, et al. Inner retinal layer thinning in radiologically isolated syndrome predicts conversion to multiple sclerosis. *Eur J Neurol*. 2020;27(11):2217-2224.
- Matute-Blanch C, Villar LM, Álvarez-Cermeño JC, et al. Neurofilament light chain and oligoclonal bands are prognostic biomarkers in radiologically isolated syndrome. *Brain*. 2018;141(4):1085-1093.
- Rossi S, Motta C, Studer V, et al. Subclinical central inflammation is risk for RIS and CIS conversion to MS. *Mult Scler*. 2015;21(11):1443-1452.
- Thouvenot E, Hinsinger G, Demattei C, et al. Cerebrospinal fluid chitinase-3-like protein 1 level is not an independent predictive factor for the risk of clinical conversion in radiologically isolated syndrome. *Mult Scler*. 2019;25(5):669-677.
- Lebrun C, Cohen M, Pignolet B, et al; on behalf SFSEP, BIONAT Network; RISC. Interleukin 17 alone is not a discriminant biomarker in early demyelinating spectrum disorders. *J Neurol Sci*. 2016;368:334-336.
- Rival M, Galoppin M, Thouvenot E. Biological markers in early multiple sclerosis: the paved way for radiologically isolated syndrome. *Front Immunol*. 2022 Apr 27;13:866092. doi: 10.3389/fimmu.2022.866092. eCollection 2022.
- Thouvenot E. Multiple sclerosis biomarkers: helping the diagnosis? *Rev Neurol (Paris)*. 2018;174(6):364-371.
- Teunissen CE, Malekzadeh A, Leurs C, Bridel C, Killestein J. Body fluid biomarkers for multiple sclerosis—the long road to clinical application. *Nat Rev Neurol*. 2015;11(10):585-596.
- Khalil M, Teunissen CE, Otto M, et al. Neurofilaments as biomarkers in neurological disorders. *Nat Rev Neurol*. 2018;14(10):577-589.
- Sellebjerg F, Royen L, Soelberg Sørensen P, Oturai AB, Jensen PEH. Prognostic value of cerebrospinal fluid neurofilament light chain and chitinase-3-like-1 in newly diagnosed patients with multiple sclerosis. *Mult Scler*. 2019;25(11):1444-1451.
- Håkansson I, Tisell A, Cassel P, et al. Neurofilament light chain in cerebrospinal fluid and prediction of disease activity in clinically isolated syndrome and relapsing-remitting multiple sclerosis. *Eur J Neurol*. 2017;24(5):703-712.
- Kuhle J, Kropshofer H, Haering DA, et al. Blood neurofilament light chain as a biomarker of MS disease activity and treatment response. *Neurology*. 2019;92(10):e1007-e1015.
- Kuhle J, Plavina T, Barro C, et al. Neurofilament light levels are associated with long-term outcomes in multiple sclerosis. *Mult Scler*. 2020;26(13):1691-1699.
- Bjornevik K, Munger KL, Cortese M, et al. Serum neurofilament light chain levels in patients with presymptomatic multiple sclerosis. *JAMA Neurol*. 2020;77(1):58-64.
- Toulouse E, Lafont B, Granier S, Mcgurk G, Bazin JE. French legal approach to patient consent in clinical research. *Anaesth Crit Care Pain Med*. 2020;39(6):883-885.
- Thompson AJ, Banwell BL, Barkhof F, et al. Diagnosis of multiple sclerosis: 2017 revisions of the McDonald criteria. *Lancet Neurol*. 2018;17(2):162-173.
- Teunissen CE, Petzold A, Bennett JL, et al. A consensus protocol for the standardization of cerebrospinal fluid collection and biobanking. *Neurology*. 2009;73(22):1914-1922.
- Thouvenot E, Demattei C, Lehmann S, et al. Serum neurofilament light chain at time of diagnosis is an independent prognostic factor of survival in amyotrophic lateral sclerosis. *Eur J Neurol*. 2020;27(2):251-257.
- Hinsinger G, Galéotti N, Nabholz N, et al. Chitinase 3-like proteins as diagnostic and prognostic biomarkers of multiple sclerosis. *Mult Scler*. 2015;21(10):1251-1261.
- Uno H, Cai T, Tian L, Wei LJ. Evaluating prediction rules for *t*-year survivors with censored regression models. *J Am Stat Assoc*. 2007;102(478):527-537.
- Uno H, Cai T, Pencina MJ, D'Agostino RB, Wei LJ. On the C-statistics for evaluating overall adequacy of risk prediction procedures with censored survival data. *Stat Med*. 2011;30(10):1105-1117.
- Gafson AR, Barthélemy NR, Bomont P, et al. Neurofilaments: neurobiological foundations for biomarker applications. *Brain*. 2020;143(7):1975-1998.
- Kuhle J, Barro C, Disanto G, et al. Serum neurofilament light chain in early relapsing remitting MS is increased and correlates with CSF levels and with MRI measures of disease severity. *Mult Scler*. 2016;22(12):1550-1559.
- Håkansson I, Tisell A, Cassel P, et al. Neurofilament levels, disease activity and brain volume during follow-up in multiple sclerosis. *J Neuroinflamm*. 2018;15(1):209.
- Gauthier A, Viel S, Perret M, et al. Comparison of Simoa and Ella to assess serum neurofilament-light chain in multiple sclerosis. *Ann Clin Transl Neurol*;8(5):1141-1150.
- Thebault S, Abdoli M, Fereshtehnejad SM, Tessier D, Tabard-Cossa V, Freedman MS. Serum neurofilament light chain predicts long term clinical outcomes in multiple sclerosis. *Sci Rep*. 2020;10(1):10381.
- Kuhle J, Barro C, Andreasson U, et al. Comparison of three analytical platforms for quantification of the neurofilament light chain in blood samples: ELISA, electrochemiluminescence immunoassay and Simoa. *Clin Chem Lab Med*. 2016;54(10):1655-1661.
- Comabella M, Fernández M, Martín R, et al. Cerebrospinal fluid chitinase 3-like 1 levels are associated with conversion to multiple sclerosis. *Brain*. 2010;133(pt 4):1082-1093.

Neurology[®] Neuroimmunology & Neuroinflammation

Neurofilament Light Chain Levels Are Predictive of Clinical Conversion in Radiologically Isolated Syndrome

Manon Rival, Eric Thouvenot, Lucile Du Trieu de Terdonck, et al.
Neurol Neuroimmunol Neuroinflamm 2023;10;
DOI 10.1212/NXI.0000000000200044

This information is current as of October 24, 2022

Updated Information & Services	including high resolution figures, can be found at: http://nn.neurology.org/content/10/1/e200044.full.html
References	This article cites 37 articles, 2 of which you can access for free at: http://nn.neurology.org/content/10/1/e200044.full.html##ref-list-1
Permissions & Licensing	Information about reproducing this article in parts (figures, tables) or in its entirety can be found online at: http://nn.neurology.org/misc/about.xhtml#permissions
Reprints	Information about ordering reprints can be found online: http://nn.neurology.org/misc/addir.xhtml#reprintsus

Neurol Neuroimmunol Neuroinflamm is an official journal of the American Academy of Neurology. Published since April 2014, it is an open-access, online-only, continuous publication journal. Copyright © 2022 The Author(s). Published by Wolters Kluwer Health, Inc. on behalf of the American Academy of Neurology. All rights reserved. Online ISSN: 2332-7812.

