Clinical/Scientific Notes

Peter Körtvelyessy, MD* Jan Bauer, PhD* Christian M. Stoppel, MD, PhD Wolfgang Brück, MD Ivonne Gerth, MA Stefan Vielhaber, MD Falk R. Wiedemann, MD Hans J. Heinze, MD Claudius Bartels, MD Christian G. Bien, MD

Neurol Neuroimmunol Neuroinflamm 2015;2:e75; doi: 10.1212/ NXI.000000000000075

COMPLEMENT-ASSOCIATED NEURONAL LOSS IN A PATIENT WITH CASPR2 ANTIBODY-ASSOCIATED ENCEPHALITIS

Recently, CASPR2 antibody–associated encephalitis was presented as a subentity of encephalitis with antibodies against the voltage-gated potassium channel (VGKC) complex. Besides limbic encephalitis, CASPR2 antibodies are also found in Isaac syndrome and Morvan syndrome.^{1,2} The specific underlying pathogenic mechanisms in limbic encephalitis are unknown. Our data suggest an antibody- and complement-mediated pathology in CASPR2 antibody–associated encephalitis.

Since 2010, our 62-year-old male patient has had unrecognized dyscognitive seizures and acoustic hallucinations. In 2012, a loss of short-term memory was also noticed. EEG and MRI conducted in August 2012 were normal (figure, A). In January 2013, his symptoms worsened and he was admitted to a hospital. At this time, MRI examination showed volume and T2/fluid-attenuated inversion recovery (FLAIR) signal increase in the right hippocampus and amygdala (figure, A).

Due to suspicion of a malignant process, the right hippocampus and parts of the temporal lobe, including the amygdala, were resected. Neuropathologic investigations ruled out tumor formation. Detailed evaluation, however, showed moderate parenchymal presence of CD3⁺ and CD8⁺ T lymphocytes (figure, C). The lack of these cells in apposition to neurons suggests the absence of T cell-mediated neuronal destruction. CD68 staining showed mild activation of microglial cells (figure, D). Furthermore, CD20 and CD138 immunostaining showed small numbers of B cells (figure, E) and plasma cells (figure, F), predominantly in the perivascular space of blood vessels. Immunoglobulin (Ig) staining showed a strong leakage of Ig through the blood-brain barrier. In areas with less blood-brain barrier leakage, however, some deposition of Ig on the membranes of neurons could be detected (figure, G). In these areas some neurons showed shrinkage and nuclear changes, suggesting degeneration (figure, G). Degenerating neurons were absent in the amygdala (figure, H),

but the hippocampus revealed few degenerating neurons (figure, I). Complement deposition, like in hippocampi of other patients with VGKC-complex encephalitis,³ was detected in a few neurons (figure, J and K). Such deposition was not found in the cortex or hippocampi of NMDAR-, Hu-, Ma2-, or GAD encephalitis patients, patients with mesial temporal lobe epilepsy with hippocampal sclerosis or Alzheimer disease, or normal controls.³

In March 2013, VGKC-complex antibodyassociated encephalitis was diagnosed and methylprednisolone (MP) treatment was started (IV followed by oral administration). This resulted in a remarkable improvement of short-term memory. To detect a potential underlying malignancy, we then performed a whole-body fluorodeoxyglucose PET, which was negative. In July 2013, 2 months after the last MP IV treatment, the patient's short-term memory problems again worsened, while attentional deficits fluctuated. MRI scans now showed a progressive cortical and hippocampal atrophy (figure, A). At this time, serum and CSF were analyzed again and were found to be positive for CASPR2 antibodies (serum and CSF obtained in April, titers 1:32,000 and 1:128, respectively, see panel B of the figure). There were no antibodies to LGI1, NMDAR, GAD65, GABA_B receptors, AMPA receptors type 1 and type 2, or glycine receptors.

The antibody index (AI, i.e., the ratio between the CSF/serum quotients of CASPR2 antibody titers and total IgG concentrations) determines whether antibodies are formed intrathecally. We considered an AI >4 as an indication of intrathecal production of the specific antibody. This conservative cutoff was used because any lower cutoff may give false positive results when titers are used.³ The values were as follows: April 2013: AI = 1.1; July: AI = 1.0; and October: AI = 2.5. Therefore, CASPR2 antibodies were produced mainly peripherally.

In July 2013, in addition to oral prednisolone, monthly cyclophosphamide infusions (15 mg/kg body weight) were started (figure, A). Thereafter, CASPR2 antibodies decreased (figure, B), neuropsychological deficits vanished, and the patient's condition stabilized. Further CSF analysis showed no pleocytosis or intrathecal IgG synthesis. MRI showed a discrete FLAIR intense signal in the left



(A) Summary of the most important changes during the disease course. Shown are consecutive cerebral fluid-attenuated inversion recovery (FLAIR) MRI scans in relation to therapeutic treatment, clinical symptoms, and CASPR2 titer. The swollen hippocampus (HC) and amygdala (AM) were resected in January 2013. Note the progressive global atrophy of the brain (from August 2012 to July 2013), which largely stopped in October 2013. The left HC, however, remained FLAIR intense in October 2013. (B) Changes in CASPR2 antibody (ab) titers in serum (dots) and CSF (squares) during the disease course. The graph shows a decline of the titers in serum and CSF after prolonged administration of cyclophosphamide and oral prednisolone (indicated by the gray bars on top). The star indicates the time point of the biopsy. (C-K) Neuropathology of the right hippocampus. (C) Double staining for CD3 and CD8 shows moderate numbers of T cells (CD8⁺ T cells are blue, CD3⁺CD8⁻ [CD4⁺] T cells are brown). (D) CD68 staining shows moderately activated microglial cells. (E) Staining for CD20 shows the presence of B cells in a perivascular cuff. (F) Staining for CD138 shows the presence of some plasma cells. (G) Staining for immuno-globulins (Ig) reveals strong leakage into the parenchyma. Deposition of immunoglobulin on neuronal membranes is indicated by the arrowheads. The arrow in the upper corner points at a degenerating neuron with nuclear changes. The inset shows an additional degenerating neuron (arrow) with a condensed nucleus. (H) Staining for TUNEL (black) and MAP2 shows the absence of degenerating cells in the amygdala of this patient. (I) In the hippocampus, a single TUNEL-positive neuron is indicated by the arrowhead. The insets show an enlargement of this neuron (left side) and a second MAP2+ TUNEL+ neuron. (J, K) Staining for complement C9neo (end complex) reveals a neuron with minor deposition, depicted by arrows (J), and a neuron with major deposition (K). CP = cyclophosphamide; MP = methylprednisolone.

Figure

© 2015 American Academy of Neurology. Unauthorized reproduction of this article is prohibited.

hippocampus but no further progression of global atrophy (figure, A). We had no indications for neuromyotonia or myasthenia gravis as CASPR2-associated symptoms at any time.

The case study presented here can refine our knowledge of CASPR2 antibody-associated encephalitis. We found signs of an antibody- and complement-mediated inflammation, which is consistent with our earlier observation in VGKCcomplex encephalitis.⁴ These findings are in contrast to NMDAR encephalitis, in which no evidence of complement deposition was found.^{5,6} This is the first description of neuronal complement activation in a patient with CASPR2 encephalitis. The features correlate well with the MRI-documented hippocampal and cerebral atrophy. In our case, cyclophosphamide combined with oral prednisolone induced a partial remission. This partial remission was, as described earlier,7 correlated with a parallel decline of serum and CSF antibodies. Therefore, clinical deficits in this condition seem to be related to 2 encephalitis-related processes: (1) a destructive, complement-mediated process leading to irreversible brain tissue loss, and (2) a direct, antibody-mediated functional process that can be stopped by removal of antibodies from CSF and serum and that results in an almost immediate clinical improvement.

* These authors contributed equally.

From the Department of Neurology (P.K., I.G., S.V., H.J.H., C.B.), University Clinic Magdeburg, Germany; Department of Neuroimmunology (J.B.), Center for Brain Research, Medical University of Vienna, Austria; Department of Psychiatry and Psychotherapy (C.M.S.), Charité University Hospital, Berlin, Germany; Department of Neuropathology (W.B.), University Medical Center Göttingen, Germany; Department of Neurology (F.R.W.), Median Clinic NRZ Magdeburg, Germany; and Epilepsy Center Bethel (C.G.B.), Krankenhaus Mara, Bielefeld, Germany.

Author contributions: P. Körtvelyessy: study concept and design, acquisition of data, analysis and interpretation, critical revision of the manuscript for important intellectual content, study supervision. J. Bauer: study concept and design, acquisition of data, analysis and interpretation, study supervision, critical revision of the manuscript for important intellectual content. C.M. Stoppel: acquisition of data, critical revision of the manuscript for important intellectual content. W. Brück: acquisition of data, critical revision of the manuscript for important intellectual content. I. Gerth: acquisition of data, analysis and interpretation. S. Vielhaber: acquisition of data, analysis and interpretation. F.R. Wiedemann: acquisition of data, study supervision. H.J. Heinze: study concept and design, study supervision. C. Bartels: acquisition of data, critical revision of the manuscript for important intellectual content. C.G. Bien: study concept and design, acquisition of data, analysis and interpretation, study supervision, critical revision of the manuscript for important intellectual content.

Study funding: No targeted funding reported.

Disclosure: P. Körtvelyessy reports no disclosures. J. Bauer is on the scientific advisory board for the Dutch Foundation for Multiple Sclerosis Research. C.M. Stoppel reports no disclosures, W. Brück serves on the advisory boards for Genzyme, Novartis, Biogen Idec Germany, and Teva Pharma; has received honoraria from Teva, Sanofi, Genzyme, Novartis, Merck-Serono, and Biogen Idec; is on the editorial board for Acta Neuropathologica, Therapeutic Advances in Neurological Disorders, Multiple Sclerosis International, and Neuropathology and Applied Neurobiology; and has received research support from the German Research Foundation, German Ministry for Science and Education, and Tschira Foundation. I. Gerth, S. Vielhaber, F.R. Wiedemann, H.J. Heinze, and C. Bartels report no disclosures. C.G. Bien is on the scientific advisory board for Eisai and UCB; has received honoraria or funding from Grifols, UCB, Eisai, GlaxoSmithKline, Destin, Biogen Idec, Fresenius Medical Care, and Diamed; has received research support from Fresenius Medical Care, Diamed, University of Bonn, Hagedorn Foundation Bielefeld, and The Krankenhaus Mara GmbH; and receives payment for antibody tests (antineural antibodies) performed in the laboratory run by C.G.B. Go to Neurology.org/nn for full disclosures. The Article Processing Charge was paid by the Department of Neurology, University of Magdeburg.

This is an open access article distributed under the terms of the Creative Commons Attribution-Noncommercial No Derivative 3.0 License, which permits downloading and sharing the work provided it is properly cited. The work cannot be changed in any way or used commercially.

Received September 19, 2014. Accepted in final form December 30, 2014.

Correspondence to Dr. Körtvelyessy: peter.koertvelyessy@med.ovgu.de

- Lancaster E, Huijbers MG, Bar V, et al. Investigations of caspr2, an autoantigen of encephalitis and neuromyotonia. Ann Neurol 2011;69:303–311.
- Irani SR, Pettingill P, Kleopa KA, et al. Antibodies to Kv1 potassium channel-complex proteins leucine-rich, glioma inactivated 1 protein and contactin-associated protein-2 in limbic encephalitis, Morvan's syndrome and acquired neuromyotonia. Brain 2010;133:2734–2748.
- Reiber H, Lange P. Quantification of virus-specific antibodies in cerebrospinal fluid and serum: sensitive and specific detection of antibody synthesis in brain. Clin Chem 1991;37:1153–1160.
- Bien CG, Vincent A, Barnett MH, et al. Immunopathology of autoantibody-associated encephalitides: clues for pathogenesis. Brain 2012;135:1622–1638.
- Martinez-Hernandez E, Horvath J, Shiloh-Malawsky Y, et al. Analysis of complement and plasma cells in the brain of patients with anti-NMDAR encephalitis. Neurology 2011;77:589–593.
- Tuzun E, Zhou L, Baehring JM, et al. Evidence for antibody-mediated pathogenesis in anti-NMDAR encephalitis associated with ovarian teratoma. Acta Neuropathol 2009;118:737–743.
- Paterson RW, Zandi MS, Armstrong R, et al. Clinical relevance of positive voltage-gated potassium channel (VGKC)complex antibodies: experience from a tertiary referral centre. J Neurol Neurosurg Psychiatry 2014;85:625–630.

Neurology[®] Neuroimmunology & Neuroinflammation

Complement-associated neuronal loss in a patient with CASPR2 antibody-associated encephalitis

Peter Körtvelyessy, Jan Bauer, Christian M. Stoppel, et al. Neurol Neuroimmunol Neuroinflamm 2015;2; DOI 10.1212/NXI.000000000000075

Updated Information & Services	including high resolution figures, can be found at: http://nn.neurology.org/content/2/2/e75.full.html
References	This article cites 7 articles, 2 of which you can access for free at: http://nn.neurology.org/content/2/2/e75.full.html##ref-list-1
Subspecialty Collections	This article, along with others on similar topics, appears in the following collection(s): All Clinical Neurology http://nn.neurology.org//cgi/collection/all_clinical_neurology All Epilepsy/Seizures http://nn.neurology.org//cgi/collection/all_epilepsy_seizures Autoimmune diseases http://nn.neurology.org//cgi/collection/autoimmune_diseases
Permissions & Licensing	Information about reproducing this article in parts (figures,tables) or in its entirety can be found online at: http://nn.neurology.org/misc/about.xhtml#permissions
Reprints	Information about ordering reprints can be found online: http://nn.neurology.org/misc/addir.xhtml#reprintsus

This information is current as of February 12, 2015

Neurol Neuroimmunol Neuroinflamm is an official journal of the American Academy of Neurology. Published since April 2014, it is an open-access, online-only, continuous publication journal. Copyright © 2015 American Academy of Neurology. All rights reserved. Online ISSN: 2332-7812.

